#### **Supplementary Information**

#### Supplementary method

# 1. Genome-wide association studies (GWAS) data of OP

The osteoporosis genome-wide association studies (GWAS) data set was obtained from the large-scale GWAS meta-analysis of osteoporosis from Genetic Factors for Osteoporosis Consortium (GEFOS, http:// www.gefos.org/).<sup>1</sup> The bone mineral density (BMD) of the femoral neck, lumbar spine, and forearm were assessed via Lunar, Hologic or Norland radiograph BMD measurements following standard procedure. BMD was adjusted for age, age-squared, gender, and weight. Whole-genome sequencing, whole-exome sequencing, and SNP array testing were performed for genotyping using commercial platforms, such as Illumina HiSeq 2000 (Illumina, Inc., San Diego, California), Illumina HumanOmni1 Quad v1-0B array (Illumina, Inc., San Diego, California), or Kompetitive Allele Specific PCR (KASP) genotyping assays (LGC, Teddington, United Kingdom). Imputation analysis was conducted by IMPUTE2 software against the combined UK10K/1000 Genomes reference panel.<sup>2-4</sup> Meta-analysis of cohortlevel common SNP association statistics was undertaken using fixed-effects meta-analysis in genome-wide association meta-analysis (GWAMA) software.<sup>5</sup> A total of 32 735 patients with genotype data were used for the GWAS of the neck of femur BMD, 28 498 for the lumbar spine, and 8143 for the forearm. A detailed description of the cohort information is in the paper by Zheng et al.<sup>1</sup>

## 2. Genome-wide expression quantitative trait loci (eQTLs) dataset

We used the recently published genome-wide eQTLs annotation map established by Westra et al.<sup>6</sup> Briefly, this eQTLs dataset was derived from a meta-analysis of 5311 samples. Gene expression profiling of blood samples was performed by Illumina whole-genome Expression BeadChips (HT12v3, HT12v4 or H8v2 arrays). SNP genotyping was evaluated by multiple chips, such as Illumina HumanHap300, HumanHap370, or the 610-Quad platforms (all Illumina, Inc., San Diego, California). Genotypes were imputed against HapMap or 1000 Genome reference panels using IMPUTE or MACH software.<sup>2-4</sup> eQTLs association tests were performed using Spearman's rank correlation, weighted for the square root of the sample size. We used a total of 923 021 ciseQTLs for 14 329 gene expression probes, identified at a false discovery rate (FDR) < 0.05. A detailed description of the study subjects and experiment design can be found in the published study by Westra et al.<sup>6</sup> The data were pre-processed by SMR software,<sup>7</sup> as described in the following section.

# 3. Genome-wide methylation quantitative trait loci (meQTLs) dataset

A genome-wide meQTLs annotation map was obtained from the latest meQTLs study of 697 Swedish individuals by McClay et al.<sup>8</sup> Genotyping was conducted using Affymetrix SNP Arrays 5.0 or 6.0(Affymetrix, California, US), or Illumina OmniExpress array (Illumina, Inc., San Diego,CA). After phasing genotypes with MACH 1.0, a 1000 Genome reference panel (phase I, version 3) was employed for imputation.<sup>2,4</sup> Genome-wide DNA methylation was evaluated using methyl-CpG binding domain (MBD) protein-based enrichment and sequencing (MBDseq) (SOLiD; Life Technologies, Thermo Fisher Scientific, Waltham, Massachusetts). Matrix eQTL was used for testing for association between genotype and methylation measurements. The final dataset comprised 683 152 unique methylation sites with local meQTLs, identified at FDR < 0.01.

### 4. Summary data-based Mendelian randomisation (SMR) analysis

SMR employed the concept of Mendelian randomisation (MR) to evaluate the effect of gene expression on traits.<sup>7</sup> The original MR analysis uses a genetic variant (*z*) as an instrumental variable to estimate and test for the causative effect of an exposure variable (*x*) on an outcome (*y*). Considering the exposure variable (*x*) to be the expression level of a gene, using a two-step least squares (2SLS) estimation, the effect of gene expression *x* on trait *y* can be expressed by  $\hat{b}_{xy} = \frac{\hat{b}_{zy}}{\hat{b}_{zx}}$ , where  $\hat{b}_{zy}$  and  $\hat{b}_{zx}$  are the least squares estimates of *y* and *x* on *z*, respectively. The sampling variance of the 2SLS estimate of  $\hat{b}_{xy}$  is  $\frac{\text{var}(y)(1-R_{xy}^2)}{n\text{var}(x)R_{xy}^2}$ . Therefore, chi-squared testing statistics can be built to test the significance of  $\hat{b}_{xy}$  based on  $\hat{b}_{xy}$  and its variance TMR =  $\frac{\hat{b}_{xy}^2}{\text{var}(\hat{b}_{xy})}$ . Compared with directly estimating  $\hat{b}_{xy}$ ,

this two-step approach provides a more accurate estimation for effect size of x on y, free of confounding from non-genetic factors.

However, such MR analysis requires genotype, gene expression, and phenotype available in the same sample, as well as the availability of individual-level data. In comparison, SMR used expression quantitative trait loci (eQTL) SNP effect data from published eQTL studies to estimate the effect of SNP on gene expression,  $\hat{b}_{zx}$ . SNP effect data from existing trait GWAS summary were used to estimate SNP effect on trait,  $\hat{b}_{zy}$ . This yielded an adjusted statistic TSMR =  $\frac{\hat{b}_{xy}^2}{\operatorname{var}(\hat{b}_{xy})} \approx \frac{z_{zy}^2 z_{zx}^2}{z_{zy}^2 + z_{zx}^2}$  where  $z_{zy}$  and  $z_{zx}$  are the z statistics from a given published GWAS and eQTL study.

 Table i.
 Expression quantitative trait loci (eQTLs)-based gene set enrichment analysis results.

Gene set name	p-value*			
	Forearm	Femoral neck	Lumbar spine	
ABE INNER EAR	4.1E-02	1.8E-02	-	
AGUIRRE PANCREATIC CANCER COPY NUMBER UP	4.1E-02	-	3.0E-02	
AMINO_SUGAR_METABOLIC_PROCESS	2.4E-02	-	4.7E-02	
AMIT_SERUM_RESPONSE_240_MCF10A	3.0E-02	-	6.0E-03	
ASTON_MAJOR_DEPRESSIVE_DISORDER_UP	1.3E-02	-	9.0E-03	
ATGTCAC,MIR-489	1.1E-02	-	1.1E-02	
BIOCARTA_P38MAPK_PATHWAY	3.9E-02	2.1E-02	-	
BROWNE_HCMV_INFECTION_24HR_DN	-	2.9E-02	6.5E-03	
CARBOXYLIC_ACID_TRANSPORT	1.7E-02	4.8E-02	-	
CASPASE_ACTIVATION	-	4.2E-02	5.0E-02	
CHIANG_LIVER_CANCER_SUBCLASS_UNANNOTATED_UP	4.0E-02	-	2.6E-02	
chr11p11	4.9E-02	-	2.5E-03	
chr11p15	4.6E-02	1.1E-02	-	
chr11q13	3.9E-02	-	1.1E-02	
chr14q31	2.1E-02	3.5E-02	-	
chr3p25	-	1.1E-02	2.5E-03	
CONDENSED_CHROMOSOME	3.3E-02	-	2.7E-02	
CTCTATG,MIR-368	-	4.7E-02	3.2E-02	
CYTOSKELETON_DEPENDENT_INTRACELLULAR_TRANSPORT	4.6E-02	-	2.6E-02	
DARWICHE_PAPILLOMA_RISK_HIGH_UP	1.0E-03	4.0E-02	3.8E-02	
DER_IFN_BETA_RESPONSE_UP	2.9E-02	-	3.7E-02	
EHLERS_ANEUPLOIDY_UP	6.0E-03	-	2.9E-02	
ESC_11_UP_EARLY.V1_UP	-	4.5E-02	4.3E-02	
GAUSSMANN_MLL_AF4_FUSION_TARGETS_A_DN	4.5E-02	-	4.0E-03	
GAVIN_FOXP3_TARGETS_CLUSTER_P7	-	1.0E-02	1.2E-02	
GCM_BMPR2	7.0E-03	-	3.6E-02	
GCM_VAV1	-	2.4E-02	1.8E-02	
GNATENKO_PLATELET_SIGNATURE	4.6E-02	-	1.0E-02	
GNF2_CD48	-	1.1E-02	1.0E-02	
UNF2_STAT6	2.0E-03	2.7E-02	-	
GSE10240_ILTZ_VS_ILTZ_AND_ILZZ_STIM_PKIMAKY_BKONCHIAL_EPTTHELIAL_CELLS_UP	1.3E-02	-	2.1E-02	
GSE12198_NK_VS_NK_ACT_EXPANSION_SYSTEM_DERIVED_NK_CELL_DN	3.7E-02	4.2E-02	-	
GSE13173_UNTREATED_VS_ILT2_TREATED_ACT_CD8_TCELL_DN	-	2.1E-02	7.0E-03	
	9.0E-03	2.2E-U2	-	
	0.0E-05	2.0E-02	2.16-02	
	- 2 3 5 0 2	7.5L-03 4.5E.02	2.72-02	
CSE17324_NAIVE_VS_ACTIVATED_LLT4_KO_CD8_TCLLL_OF	2.3L-02	4.5L-02 1 /F-02	-	
CSE17700_WILMONI_VS_INAIVL_DCLLL_OF	2.7 L-02	1.4L-02 4 7E-02	- 3 4E-02	
CSE17721_ETRE_VS_TANDESR4_T2T_DMDM_UP	_	3 4F-02	1 3E-02	
CSE17721_ES_VS_POLVIC_4H_BMDM_UP	- 1 5E-02	5.42-02	3 0F-02	
GSE17721_EIS_VS_IGEIRE_HI_BMDM_OF	3.8E-02	2 8F-02	5.0E-02	
GSE17721_PAM3CSK4_VS_CPG_16H_BMDM_UP	2 3E-02	-	2 1F-02	
GSE17721 POLYIC VS CPG 1H BMDM DN	3.1F-02	-	1.5E-02	
GSE17721 POLYIC VS PAM3CSK4 8H BMDM DN	4.0E-02	4.1E-02	-	
GSE17974 CTRL VS ACT IL4 AND ANTI IL12 1H CD4 TCELL UP	-	2.3E-02	2.7E-02	
GSE19888 CTRL VS T CELL MEMBRANES ACT MAST CELL UP	8.5E-03	-	1.7E-02	
GSE20152 HTNFA OVERXPRESS ANKLE VS CTRL SPHK1 KO ANKLE UP	3.0E-03	-	4.5E-02	
GSE21033 CTRL VS POLYIC STIM DC 1H DN	6.0E-03	2.1E-02	-	
GSE21380 TFH VS GERMINAL CENTER TFH CD4 TCELL DN	2.4E-02	-	2.3E-02	
GSE22589_HEALTHY_VS_SIV_INFECTED_DC_UP	-	2.0E-03	4.3E-02	
GSE22611_NOD2_VS_MUTANT_NOD2_TRANSDUCED_HEK293T_CELL_UP	3.0E-02	-	3.1E-02	
GSE22886_UNSTIM_VS_IL15_STIM_NKCELL_UP	-	2.1E-02	2.1E-02	
GSE24574_BCL6_HIGH_TFH_VS_TFH_CD4_TCELL_UP	4.2E-02	-	2.2E-02	
GSE24634_TREG_VS_TCONV_POST_DAY3_IL4_CONVERSION_UP	4.5E-02	4.5E-02	-	
GSE2585_THYMIC_DC_VS_THYMIC_MACROPHAGE_UP	3.4E-02	-	4.4E-02	
GSE2770_IL4_ACT_VS_ACT_CD4_TCELL_6H_UP	2.3E-02	2.0E-03	-	
GSE2770_UNTREATED_VS_TGFB_AND_IL12_TREATED_ACT_CD4_TCELL_6H_UP	-	1.4E-02	7.5E-03	
GSE27786_LIN_NEG_VS_ERYTHROBLAST_DN	-	2.5E-03	2.6E-02	
GSE29949_MICROGLIA_VS_DC_BRAIN_DN	3.3E-02	4.2E-02	-	
GSE31082_DP_VS_CD4_SP_THYMOCYTE_DN	4.3E-02	-	6.5E-03	
GSE31082_DP_VS_CD8_SP_THYMOCYTE_DN	3.2E-02	-	4.9E-02	
GSE32034_LY6C_HIGH_VS_LOW_ROSIGLIZATONE_TREATED_MONOCYTE_UP	4.0E-02	2.5E-02	-	
GSE32164_RESTING_DIFFERENTIATED_VS_CMYC_INHIBITED_MACROPHAGE_UP	1.3E-02	-	3.6E-02	
GSE33292_DN3_THYMOCYTE_VS_TCF1_KO_TCELL_LYMPHOMA_UP	-	1.0E-03	4.0E-02	
GSE33424_CD161_HIGH_VS_INT_CD8_TCELL_UP	3.0E-02	-	8.5E-03	

(Continued)

#### Table i. (Continued)

Gene set name		p-value*			
	Forearm	Femoral neck	Lumbar spine		
GSE34217_MIR17_92_OVEREXPRESS_VS_WT_ACT_CD8_TCELL_UP	2.3E-02	-	3.7E-02		
GSE360_CTRL_VS_L_MAJOR_DC_DN	-	3.7E-02	1.1E-02		
GSE369_PRE_VS_POST_IL6_INJECTION_SOCS3_KO_LIVER_UP	1.8E-02	-	3.5E-02		
GSE37301_COMMON_LYMPHOID_PROGENITOR_VS_CD4_TCELL_UP	-	3.8E-02	4.9E-02		
GSE37301_LYMPHOID_PRIMED_MPP_VS_COMMON_LYMPHOID_PROGENITOR_DN	2.0E-03	5.0E-03	-		
GSE37301_MULTIPOTENT_PROGENITOR_VS_GRAN_MONO_PROGENITOR_UP	3.5E-03	5.0E-04	-		
GSE37563_WT_VS_CTLA4_KO_CD4_TCELL_D4_POST_IMMUNIZATION_UP	1.0E-02	-	1.5E-02		
GSE39556_CD8A_DC_VS_NK_CELL_MOUSE_3H_POST_POLYIC_INJ_UP	4.8E-02	1.3E-02	-		
GSE39556_UNTREATED_VS_3H_POLYIC_INJ_MOUSE_CD8A_DC_DN	2.3E-02	2.7E-02	-		
GSE3982_DC_VS_NEUTROPHIL_DN	2.4E-02	-	1.1E-02		
GSE3982_EOSINOPHIL_VS_BASOPHIL_UP	1.2E-02	3.0E-03	-		
	3.3E-02	-	3.3E-UZ		
GSE3962_MAC_VS_CENT_MEMORT_CD4_TCELL_DN	4.0E-03	-	2.6E-02		
GESPOZ_MASI_CELL_VS_DC_DN	2.0E-02	1.JE-02	-		
	4.7E-02	-	4.0E-02 0.0E.03		
CSE42021_CD24E0_TREG_V3_CD24E0_TCONV_TITINIO3_OF	1.1E.02	-	9.0L-03		
CSE43700_ONTREATED_V3_IETO_TREATED_FBMC_OF	2 6E 02	-	1.50-02		
CSE43863 TH1 VS TEH MEMORY CDA TCELL LIP	1.0E-02		1.5L-02 2.4E-02		
CSE43055 1H VS 60H ACT CDA TCELL UP	1.02-02	- 3.8F-02	2.4L-02 3.3E-02		
CSE43955_TCER II & VS TCER II & II 23 TH17 ACT CD4 TCELL 52H DN	_	2 55-02	1.9E-02		
CSE43955_TOTU_LC_V3_TOTU_LC_LC25_TTTY_ACT_CCL4_TCLL_52T_DTV	- 1 2E-02	2.52-02	4 9F-02		
CSE45355_WT_V5_SORT_KO_THTY_DITTERED FOR THE REPORT OF THE	4 4F-02	- 2 1F-02	-		
CSE45365 WT VS IENAR KO CD118 DC MCMV INFECTION UP	2 3E-02	1 2F-02			
CSE46606 IREA KO VS WT LINSTIM RCELL DN	2.3L-02 2.7E-02	1.22-02	- 9 5E-03		
CSE40000_INI 4_KO_V3_W1_0N3 IIWI_DCELE_DN	2.7 L-02	- 1 6F-02	1 2E-02		
CSESS89 ILE KO VS ILIO KO LOS AND ILIO STIM MACOOPHACE ASMIN LID	- 4 3E-02	3 65-02	2 OF-03		
CSESSOS_LEG_KO_VS_LETO_KO_LI S_XIVD_LETO_STIM_MAKENOTTIAGE_4SMINT_OT	4.5E-02 4.5E-03	5.02-02	2.0E-03		
GSE6259 BCELL VS CD8 TCELL UP	2 6F-02	_	4 5F-03		
GSE6674 LINSTIM VS ANTLIGM STIM BCELL LIP	2.0E-02 2.8F-02	_	1 OF-03		
GSE7509 DC VS MONOCYTE DN	4.9F-02	-	1.0E-03		
GSE7509 UNSTIM VS TNEA II 18 II 6 PGE STIM DC DN	3.4F-02	-	3.9F-02		
GSE7768 OVA ALONE VS OVA WITH MPL IMMUNIZED MOUSE WHOLE SPLEEN 6H UP	6.0E-03	8.5E-03	-		
GSE7852 LN VS FAT TREG DN	2.4E-02	2.9E-02	-		
GSE8621 UNSTIM VS LPS STIM MACROPHAGE DN	-	3.7E-02	3.1E-02		
GSE8685 IL2 STARVED VS IL21 ACT IL2 STARVED CD4 TCELL UP	2.4E-02	2.0E-02	-		
GSE8921 3H VS 24H TLR1 2 STIM MONOCYTE UP	-	4.5E-03	4.2E-02		
GSE8921_UNSTIM_VS_TLR1_2_STIM_MONOCYTE_24H_DN	-	2.7E-02	6.0E-03		
GSE9037_WT_VS_IRAK4_KO_LPS_4H_STIM_BMDM_DN	1.0E-03	-	6.0E-03		
GSE9988_ANTI_TREM1_VS_ANTI_TREM1_AND_LPS_MONOCYTE_DN	4.2E-02	1.5E-02	-		
GSE9988_LPS_VS_VEHICLE_TREATED_MONOCYTE_UP	3.5E-02	4.0E-02	-		
GUO_HEX_TARGETS_DN	4.5E-02	4.7E-02	2.4E-02		
HALLMARK_UV_RESPONSE_UP	4.9E-02	-	3.7E-02		
HU_GENOTOXIC_DAMAGE_24HR	4.8E-02	1.0E-02	-		
HYDROLASE_ACTIVITY_ACTING_ON_ACID_ANHYDRIDES	2.0E-02	-	9.0E-03		
HYDROLASE_ACTIVITY_ACTING_ON_CARBON_NITROGEN_NOT_PEPTIDEBONDS	-	4.5E-03	4.1E-02		
HYDROLASE_ACTIVITY_ACTING_ON_CARBON_NITROGEN_NOT_PEPTIDEBONDSIN_LINEAR_	-	9.0E-03	5.0E-04		
AMIDES					
INFLAMMATORY_RESPONSE	1.7E-02	2.7E-02	-		
INTRACELLULAR_TRANSPORT	-	3./E-02	4.1E-02		
KEGG_FOCAL_ADHESION	2.5E-03	-	3.2E-02		
KEGG_MAPK_SIGNALING_PATHWAY	1.5E-02	4./E-02	-		
KEGG_PATHWAYS_IN_CANCER	3.8E-02	4.9E-02	8.0E-03		
	5.0E-02	4.5E-02	-		
	2.5E-02	2.5E-02	-		
	1.1E-02	3.9E-02	-		
LEE_LIVER_CANCER_EZFI_DIN	4.4E-02	2.1E-02	-		
	2.0E-02	-	1.7E-02		
	3.0E-UZ	-	2.4E-02		
	5.4E-0Z	3.4E-UZ	-		
	4.3E-UZ	3.2E-UZ	-		
IVIICROTODOLL_DAJED_PROCEJJ MIKKELSENI MEE LOD WITH H2K27ME2	1.2E-UZ	-	4.3E-UZ		
	2.0E-02	-	5.0E-UZ		
	3.0E-04 1.1E-02	- 1 2E 0.2	0.JE-UJ 1 7E-02		
	5 OF-04	4.2L-02 3.9E-02	3.05-03		
MODULE 289	-	1.9F-02	3.2F-02		
			J.22 V2		

(Continued)

## Table i. (Continued)

Gene set name	p-value*		
	Forearm	Femoral neck	Lumbar spine
MODULE_334	1.0E-03	-	4.0E-03
MODULE_356	1.7E-02	-	3.2E-02
MODULE_427	1.0E-03	4.7E-02	3.0E-03
MODULE_480	5.0E-03	1.4E-02	1.4E-02
MODULE_97	4.4E-02	2.3E-02	-
MORF_DCC	-	2.7E-02	2.7E-02
NABA_ECM_GLYCOPROTEINS	4.1E-02	5.0E-02	2.8E-02
NIKOLSKY_BREAST_CANCER_16P13_AMPLICON	1.2E-02	5.0E-03	-
NUCLEOSIDE_TRIPHOSPHATASE_ACTIVITY	2.8E-02	-	1.2E-02
ORGANIC_ACID_TRANSPORT	1.7E-02	4.8E-02	-
PARK_TRETINOIN_RESPONSE_AND_RARA_PLZF_FUSION	1.0E-02	-	2.7E-02
PASINI_SUZ12_TARGETS_UP	2.5E-02	-	1.1E-02
PID_AR_NONGENOMIC_PATHWAY	3.2E-02	-	4.8E-02
PID_ECADHERIN_STABILIZATION_PATHWAY	3.3E-02	-	4.5E-02
PID_SYNDECAN_1_PATHWAY	2.3E-02	-	3.2E-02
PYROPHOSPHATASE_ACTIVITY	2.5E-02	-	2.0E-02
REACTOME ACTIVATION OF NF KAPPAB IN B CELLS	-	3.6E-02	4.8E-02
REACTOME CIRCADIAN CLOCK	-	2.7E-02	1.0E-03
REACTOME MAPK TARGETS NUCLEAR EVENTS MEDIATED BY MAP KINASES	7.0E-03	2.1E-02	-
REACTOME METABOLISM OF CARBOHYDRATES	4.2E-02	9.5E-03	-
REACTOME MYOGENESIS	8.0E-03	1.7E-02	-
REACTOME NCAM SIGNALING FOR NEURITE OUT GROWTH	1.9E-02	-	2.8E-02
REACTOME NEURONAL SYSTEM	3.4E-02	-	3.1E-02
REACTOME NUCLEAR EVENTS KINASE AND TRANSCRIPTION FACTOR ACTIVATION	6.0E-03	3.5E-02	-
REACTOME RORA ACTIVATES CIRCADIAN EXPRESSION	-	1.1E-02	5.0E-03
REACTOME_TRAF6_MEDIATED_INDUCTION_OF_NFKB_AND_MAP_KINASES_UPON_TLR7_8_ OR_9_ACTIVATION	2.0E-03	3.2E-02	-
REACTOME_TRANSMEMBRANE_TRANSPORT_OF_SMALL_MOLECULES	4.6E-02	3.0E-03	-
REACTOME_TRANSMISSION_ACROSS_CHEMICAL_SYNAPSES	8.5E-03	-	1.3E-02
REACTOME_TRIF_MEDIATED_TLR3_SIGNALING	1.6E-02	3.2E-02	-
RESPONSE_TO_CHEMICAL_STIMULUS	-	3.3E-02	4.2E-02
ROVERSI_GLIOMA_COPY_NUMBER_UP	3.0E-03	-	2.0E-03
SARTIPY_BLUNTED_BY_INSULIN_RESISTANCE_DN	2.2E-02	-	4.1E-02
SEQUENCE_SPECIFIC_DNA_BINDING	-	9.5E-03	4.5E-03
TGACATY_UNKNOWN	2.7E-02	-	3.6E-02
TGGNNNNNKCCAR_UNKNOWN	4.5E-03	-	1.6E-02
TSENG_IRS1_TARGETS_UP	-	2.5E-02	2.3E-02
V\$CRX_Q4	3.2E-02	-	3.3E-02
V\$GATA1_04	5.0E-03	-	4.7E-02
V\$HNF4_01	2.5E-03	-	2.9E-02
V\$SREBP1_Q6	4.2E-02	-	5.0E-04
V\$SRF_Q5_01	2.3E-02	-	3.8E-02
V\$TAL1BETAE47_01	1.9E-02	3.5E-03	-
V\$TCF4_Q5	3.0E-02	-	2.5E-02
V\$TEF1_Q6	4.5E-02	-	1.4E-02
VALK_AML_WITH_EVI1	3.1E-02	-	4.4E-02
WEIGEL_OXIDATIVE_STRESS_BY_HNE_AND_TBH	3.2E-02	-	5.0E-03
WELCH_GATA1_TARGETS	8.5E-03	-	1.5E-02
YAGI_AML_FAB_MARKERS	3.7E-02	8.0E-03	-
YTTCCNNNGGAMR_UNKNOWN	2.0E-03	3.7E-02	3.2E-02

\*Kolmogorov-Smirnov running sum statistics was used and p-values were decided based on permutation<sup>9</sup>

 Table ii.
 Methylation quantitative trait loci (meQTLs)-based gene set enrichment analysis results.

Gene set name	p-value'		
	Forearm	Femoral neck	Lumbar spine
ACAACCT,MIR-453	-	3.0E-03	3.1E-02
ANDROGEN_RECEPTOR_SIGNALING_PATHWAY	4.7E-02	-	3.4E-02
ANION_TRANSPORT	9.5E-03	-	1.1E-02
BIOCARTA_CLASSIC_PATHWAY	1.4E-02	-	2.8E-02
BIOCARTA_IL1R_PATHWAY	-	5.0E-02	2.7E-02
BIOCARTA_LAIR_PATHWAY	-	1.0E-02	1.6E-02
BRUECKNER_TARGETS_OF_MIRLET7A3_DN	2.8E-02	-	3.7E-02
CHANG_CYCLING_GENES	3.8E-02	1.0E-02	9.5E-03
chr6q14	2.6E-02	6.5E-03	3.0E-02
chr8p23	4.9E-02	-	5.0E-04
CTCTAGA,MIR-526C,MIR-518F,MIR-526A	2.6E-02	3.8E-02	-
DEMAGALHAES_AGING_UP	4.9E-02	4.2E-02	-
DIGESTION	-	4.6E-02	9.5E-03
	2.3E-02	-	4.6E-02
	-	2.0E-02	2.3E-02
ESTABLISHMENT_OF_ORGANELLE_LOCALIZATION	3.5E-02	1.5E-02	-
	1.1E-02	-	2.5E-02
	-	1.4E-02	4.0E-02
	2.3E-02	2.3E-03	-
	2.3E-02	-	1.2E-02
	2.4E-02 3.7E-02	4.3E-03	-
	3.7E-02	1.3E-02 4 0E 02	- 1 3E 02
	4.3E-02	4.9E-02	1.3E-02
	3.2E-02	4.3E-02 3.2E-02	1.3E-02
CNE2_PENA	4.0L-03	3.2L-02	-
	2.0L-02 4.0E-03	4.0L-02 2.2E-02	-
CNF2_TTK	3.5E-02	1 1F-02	
CSE11057 NAIVE VS CENT MEMORY CD4 TCELL DN	4 1F-02	3 5E-02	_
CSE14769 LINSTIM VS 240MIN LPS BMDM LIP	3.0E-02	5.52-02	1 3E-02
GSE15330 WT VS IKAROS KO GRANULOCYTE MONOCYTE PROGENITOR LIP	-	9 0F-03	1.9E-02
GSE17974 CTRL VS ACT IL4 AND ANTI IL12 0.5H CD4 TCELL UP	1.9E-02	2.8E-02	-
GSE18893 CTRL VS TNF TREATED TREG 24H DN	3.0E-02	-	1.5E-02
GSE1925 CTRL VS IFNG PRIMED MACROPHAGE 3H IFNG STIM DN	-	4.5E-03	4.4E-02
GSE19772 CTRL VS HCMV INF MONOCYTES UP	-	9.5E-03	2.3E-02
GSE21063 3H VS 16H ANTI IGM STIM BCELL DN	3.6E-02	-	3.7E-02
GSE21063_WT_VS_NFATC1_KO_8H_ANTI_IGM_STIM_BCELL_UP	3.0E-03	2.5E-03	-
GSE21379_TFH_VS_NON_TFH_CD4_TCELL_UP	2.2E-02	-	1.9E-02
GSE22886_CD8_VS_CD4_NAIVE_TCELL_DN	-	2.0E-03	4.0E-02
GSE23925_DARK_ZONE_VS_NAIVE_BCELL_DN	1.9E-02	1.3E-02	-
GSE25085_FETAL_LIVER_VS_ADULT_BM_SP4_THYMIC_IMPLANT_DN	1.9E-02	3.1E-02	-
GSE25088_CTRL_VS_ROSIGLITAZONE_STIM_MACROPHAGE_UP	-	1.0E-02	3.0E-03
GSE25123_CTRL_VS_IL4_STIM_MACROPHAGE_UP	-	3.1E-02	2.0E-03
GSE2585_CTEC_VS_MTEC_THYMUS_UP	4.1E-02	4.0E-02	-
GSE2826_WT_VS_XID_BCELL_DN	-	7.5E-03	3.0E-02
GSE29614_CTRL_VS_DAY3_TIV_FLU_VACCINE_PBMC_DN	2.1E-02	-	4.8E-02
GSE29618_PRE_VS_DAY7_POST_TIV_FLU_VACCINE_PDC_DN	3.1E-02	1.2E-02	-
GSE30962_PRIMARY_VS_SECONDARY_ACUTE_LCMV_INF_CD8_TCELL_UP	1.1E-02	3.3E-02	-
GSE36392_EOSINOPHIL_VS_MAC_IL25_TREATED_LUNG_UP	-	7.5E-03	5.0E-02
GSE3982_MAST_CELL_VS_NKCELL_DN	-	1.1E-02	1.5E-02
GSE41867_DAY8_VS_DAY15_LCMV_ARMSTRONG_EFFECTOR_CD8_TCELL_UP	1.7E-02	-	1.8E-02
GSE41867_NAIVE_VS_DAY8_LCMV_CLONE13_EFFECTOR_CD8_TCELL_DN	1./E-02	2.5E-02	-
GSE42088_2H_VS_24H_LEISHMANIA_INF_DC_DN	-	1.4E-02	2.5E-02
GSE43863_DAY6_EFF_VS_DAY150_MEM_LY6C_IN1_CXCR5POS_CD4_ICELL_DN	-	2.5E-03	2.5E-02
GSE43863_NAIVE_VS_LY6C_INT_CXCK5PUS_CD4_EFF_TCELL_D6_LCMV_DN	7.0E-03	-	4.0E-02
GSE43955_IH_V5_60H_ACI_CD4_ICELL_UP	-	1.9E-02	2.4E-02
	-	3.0E-02	3.0E-02
	-	3.0E-02	4.0E-02
GSE/768_OVA_WITH_LPS_VS_OVA_WITH_MPL_IMMUNIZED_MOUSE_WHOLE_SPLEEN_6H_UP	3.0E-02	3.0E-02	-
	-	4.0E-UZ	5.5E-0Z
	1.3E-UZ	3.7E-UZ	-
	3.UE-UZ	4.3E-U3	-
	3.UE-UZ	5.3E-U3	- 2 6E 02
	-	3.UE-U4 2.1E-02	2.0E-UZ
	1.72-02	2.1E-UZ 8 0E 02	-
KARI SSON TGERT TARGETS UP	1.0L-02 1.4F-02	0.01-03	- 4 9F-02
		-	

## Table ii. (Continued)

Gene set name	p-value*		
	Forearm	Femoral neck	Lumbar spine
KEGG LEUKOCYTE TRANSENDOTHELIAL MIGRATION	2.3E-02	6.0E-03	-
KOKKINAKIS METHIONINE DEPRIVATION 96HR DN	1.4E-02	-	1.0E-02
KORKOLA YOLK SAC TUMOR UP	4.9E-02	-	4.5E-02
KRAS.AMP.LUNG UP.V1 UP	_	4.8E-02	3.6E-02
LEE AGING NEOCORTEX DN	4.5E-02	-	4.0E-03
LENAOUR DENDRITIC CELL MATURATION DN	4.7F-02	_	4.4F-02
	-	4.9F-02	4.0F-02
MODILLE 147	2 7F-02	6 0E-03	-
MODULE 256	3.4F-02	5.0E-03	_
MODULE 451	5.42-02	4.7E-02	2 0F-02
	- 4 4E-02	5.0E-04	2.02-02
MYLLYKANCAS AMPLIEICATION HOT SPOT 12	7.72-02	2 05 02	- 2 7E 02
	-	2.91-02	2.7L-02 4.6E.02
	- 2 7E 0.2	5.5E-02 1.2E.02	4.0E-02
	5.7E-02	1.2E-02	-
ORGANELLE_LOCALIZATION	-	2.5E-03	3.0E-02
OSWALD_HEMATOPOIETIC_STEM_CELL_IN_COLLAGEN_GEL_DN	-	4.2E-02	3.6E-02
PROTEIN_COMPLEX_BINDING	3.5E-03	3.3E-02	-
PUJANA_BREASI_CANCER_LII_INI_NETWORK	1.0E-02	-	3.5E-02
RADAEVA_RESPONSE_TO_IFNA1_UP	3.6E-02	-	4.4E-02
RAMPON_ENRICHED_LEARNING_ENVIRONMENT_LATE_UP	-	5.0E-02	2.6E-02
REACTOME_AMINE_COMPOUND_SLC_TRANSPORTERS	-	4.6E-02	2.8E-02
REACTOME_FANCONI_ANEMIA_PATHWAY	-	6.0E-03	2.8E-02
REACTOME_GROWTH_HORMONE_RECEPTOR_SIGNALING	-	1.5E-02	4.4E-02
REACTOME_HYALURONAN_METABOLISM	6.0E-03	-	2.0E-03
REACTOME_HYALURONAN_UPTAKE_AND_DEGRADATION	1.6E-02	-	3.0E-03
REACTOME_LOSS_OF_NLP_FROM_MITOTIC_CENTROSOMES	5.5E-03	7.5E-03	-
REACTOME_RECRUITMENT_OF_MITOTIC_CENTROSOME_PROTEINS_AND_COMPLEXES	4.0E-03	1.1E-02	-
REACTOME_TRANSPORT_OF_VITAMINS_NUCLEOSIDES_AND_RELATED_MOLECULES	-	1.3E-02	1.0E-03
RODRIGUES_NTN1_AND_DCC_TARGETS	-	4.2E-02	3.4E-02
SPINDLE_POLE	4.6E-02	-	4.9E-02
SPIRA SMOKERS LUNG CANCER UP	-	4.8E-02	4.1E-02
TRANSCRIPTION COACTIVATOR ACTIVITY	-	4.4E-02	3.1E-02
UROSEVIC RESPONSE TO IMIQUIMOD	-	4.9E-02	4.8E-02
V\$CREB O4	-	3.6E-02	4.5E-03
V\$E2F 03	1.3E-02	-	1.4E-02
V\$F2F_O3	4.0F-03	-	2.0F-03
V\$E2E_Q3	7 OF-03	_	1 5E-03
V\$E2E_Q3_01	1 7F-02	_	3 OF-03
V\$E2E1_03	1.7 E 02 1.3E-02	4 9F-02	5.02 05
V\$E2E1_Q3	5.0E-03		1 5E-03
V\$FREAC7_01	3 SE-03	3 4E-02	-
VALK AML CLUSTER 6	3.3L-03	9.4L-02 8.0E.02	-
VEDECCUIA DESDONISE TO TCEDI CA	Z.ZE-UZ	0.UE-U3	- 9 5E 02
	3.3E-U3	-	0.JE-U3
	9.5E-U3	1.0E-UZ	-
	4./E-02	1.1E-02	-
ZHANG_ANTIVIKAL_KESPONSE_TO_KIBAVIKIN_UP	-	4.1E-02	4.6E-02
ZHANG_TLX_TARGETS_60HR_DN	-	4.5E-02	4.4E-02

\*Kolmogorov-Smirnov running sum statistics was used and p-values were decided based on permutation<sup>9</sup>

#### References

- Zheng HF, Forgetta V, Hsu YH, et al. Whole-genome sequencing identifies EN1 as a determinant of bone density and fracture. *Nature* 2015;526:112-117.
- Howie BN, Donnelly P, Marchini J. A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. *PLoS Genetics* 2009;5: e1000529.
- **3.** No authors listed. UK10K. Rare genetic variants in health and disease. What is UK10K? https://www.uk10k.org/ (date last accessed 18 September 2017).
- The 1000 Genomes Project Consortium. An integrated map of genetic variation from 1,092 human genomes. *Nature* 2012;491:56-65.
- Magi R, Morris AP. GWAMA: software for genome-wide association meta-analysis. BMC Bioinformatics 2010;11:288.
- Westra HJ, Peters MJ, Esko T, et al. Systematic identification of trans eQTLs as putative drivers of known disease associations. *Nat Genet* 2013;45: 1238-1243.
- Zhu Z, Zhang F, Hu H, et al. Integration of summary data from GWAS and eQTL studies predicts complex trait gene targets. *Nat Genet* 2016;48:481-487.
- McClay JL, Shabalin AA, Dozmorov MG, et al. High density methylation QTL analysis in human blood via next-generation sequencing of the methylated genomic DNA fraction. *Genome Biol* 2015;16:291.
- Wang K, Li M, Bucan M. Pathway-based approaches for analysis of genome wide association studies. Am J Hum Genet 2007;81:1278-1283.